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Glutamate receptor dysfunction and drug targets across models of autism spectrum disorders

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ABSTRACT

There is strong evidence that metabotropic and ionotropic glutamate receptors are affected in autism spectrum disorders (ASD), but there are few candidate genes indicating involvement of these receptors. This suggests that glutamate receptor dysregulation may primarily be involved in the expression of ASD, but is an uncommon etiology. Directly implicated in models of fragile-X with ASD phenotypes is metabotropic glutamate receptor type 5 (mGluR5), which appears to be an effective pharmacologic target in a number of models of ASD. The review of other ASD models demonstrates that there is also evidence of a role for kainate, NMDA, and AMPA receptors in the neuropathophysiology of ASD, though the relationship between dysfunction in those receptors and ASD-associated phenotypes is not well understood. Current models indicate a way forward to delineate the role of glutamate receptors in ASD. Further development of preclinical models focusing on glutamate receptors may provide tools to target a clinically important subset of ASD symptoms.

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1. Introduction

A diagnosis of ASD is determined from the presence of core deficits in 1) communication, 2) social, and 3) repetitive behaviors; with changes in social behavior and inappropriate repetitive activity being the most easily measured in rodent models (Brodtkin, 2008; Crawley, 2007; Korff and Harvey, 2006). ASD is also associated with clinically important common co-morbidities, which include anxiety, epilepsy, and intellectual disabilities. In the context of ASD, development of treatments over and within each of these domains has been hampered by the paucity of validated models of ASD.

Glutamate is the most common neurotransmitter, mediating fast-excitatory transmission in mammalian systems via AMPA and Kainate receptors (Lodge, 2009). At NMDA and metabotropic glutamate receptors (mGluR), glutamate also engages intracellular transduction mechanisms in neurons and glia to provide a broad regulatory impact on neuronal homeostasis, local and nuclear transcription, synaptic plasticity, and neuronal development. Disruption in the balance between excitation and inhibition is a commonly proposed disease mechanism in ASD, as well as in the common ASD co-morbidity of epilepsy (Orekhova et al., 2007; Polleux and Lauder, 2004). Glutamatergic synaptic transmission through AMPA receptors provides the excitatory pole of this balance. Linking NMDA and metabotropic receptors to AMPA receptor activity are a number of synaptic

plasticity mechanisms, where glutamate-evoked intracellular signaling modulates AMPA-dependent excitatory strength, via regulation of AMPA receptor cycling into the synaptic membrane (Bruneau et al., 2009; Svitkina et al., 2010). Reduced AMPA receptor density is associated with ASD in human patients (Purcell et al., 2001).

Perhaps surprisingly, there is only one strong glutamate receptor gene candidate for ASD in the kainate receptor family. (Jamain et al., 2002; Shuang et al., 2004; Strutz-Seebohm et al., 2006). However, as discussed below, glutamate transmission abnormalities are common in models of ASD. How specific glutamatergic dysfunction may lead to specific core- and associated-symptoms of ASD is not fully understood, nor are the therapeutic rationales for targeting glutamatergic systems well developed. Thus, in the face of limited effective treatments for ASD, additional tools to research the role of glutamate receptors in ASD are needed.

To review the present utility of models to enlighten the role of the glutamate system in ASD, we will first discuss preclinical models designed with transgenic construct validity for disruption of glutamatergic transmission. The balance of our discussion reviews models that recapitulate features of ASD that may in turn be relevant for delineating a relationship between ASD related phenotypes and glutamate receptor dysfunction. Finally, we briefly speculate on new directions in model development targeted at the glutamatergic system in ASD.

2. Modeling genetic candidates of ASD: GluR6

ASD candidate genes, coding for proteins directly involved in glutamatergic transmission, are rare, yet a subunit of the Kainate receptor GluR6 was identified as a strong candidate gene (Jamain et

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al., 2002; Shuang et al., 2004; Strutz-Seeböhm et al., 2006). These findings were most recently replicated in European- and Korean-derived populations (Holt et al., 2010; Kim et al., 2007). Along with AMPA receptors, kainate receptors make up the fast-ionotropic glutamate receptors and are differentiated into subtypes by the presence of GluR5 or GluR6. While a minor contributor overall, kainate receptors provide important sources of excitatory drive, particularly in interneurons, and they are also implicated in some forms of plasticity (Fisahn et al., 2004; Kullmann and Lamsa, 2008; Rodriguez-Moreno and Sihra, 2007). Kainate receptors can also have metabotropic activity that may mediate some of its actions (Fisahn et al., 2005; Rodriguez-Moreno and Sihra, 2007). GluR6-containing receptors also strongly modulate gamma-band EEG activity, an area of growing interest clinically and preclinically in ASD (Fisahn et al., 2004; Gandal et al., 2010; Rojas et al., 2008; Wilson et al., 2007). Mice without GluR6 showed reduced susceptibility to seizures and appeared less anxious, indicating that GluR6 may have a role in ASD co-morbid symptom domains of anxiety and epilepsy (Shaltiel et al., 2008). One of the polymorphisms linked to ASD as well as to obsessive-compulsive disorder (OCD; M8361) appears to produce a gain of function, with this polymorphism generating larger kainate-induced currents (Shuang et al., 2004). In contrast to the GluR6 KO mice, such a gain of function could be predicted to be anxiogenic, affect gamma-band activity, and increase seizure susceptibility and OCD-like behavioral correlates. Thus, the generation of mice carrying a knocked-in M8361 allele may provide a useful tool for preclinical drug development targeting abnormal GluR6-mediated activity in ASD.

3. Fragile-X, FMR1 and the mGluR5 hypothesis of ASD

Fragile X is a leading cause of inherited intellectual disabilities and is associated with the presence of various social disabilities that, in some cases, lead to diagnoses of ASD. Models developed in a number of systems to understand and test mechanisms of fragile-X are exemplary for identifying a role for metabotropic glutamate receptors (mGluR) in ASD (Bear et al., 2004). Fragile X was identified as a chromosomal abnormality with a number of genes potentially involved, but the specific disruption of the gene FMR1 (coding for the fragile-X mental retardation protein 1) has been strongly implicated in the syndrome (Kaytor and Orr, 2001). Nearly three decades of research have generated a strong molecular, neurodevelopmental and synaptic story that describes how reduced FMR1 leads to a loss of mRNA translational repression, allowing an upregulation of type 5 metabotropic glutamate receptor (mGluR5) activity (Bear et al., 2004; Kaytor and Orr, 2001). This increased mGluR5 activity underlies some of the symptomatology of fragile-X in model systems (Westmark et al., 2009; Yan et al., 2005; but see Dahlhaus and El-Husseini, 2010). This work has advanced to clinical trials with mGluR5 antagonists in fragile-X patients (Dolen et al., 2010).

Much of this mGluR5 hypothesis of fragile-X was established by disrupting FMR1 in mice, zebrafish and *Drosophila* (Bardoni and Mandel, 2002; den Broeder et al., 2009; McBride et al., 2005; Morales et al., 2002; Tucker et al., 2006; Wan et al., 2000). These models identified the molecular mechanism of FMR1 as a local suppressor of mRNA translation, leading to altered dendritic morphology, reduced excitatory potentials, and disrupted synaptic plasticity (well reviewed in (Bear et al., 2004; Dolen and Bear, 2008). On the behavioral level, face validity for core features of ASD, mice and *Drosophila* models with deleted FMR1 showed impaired social interactions (McBride et al., 2005; McNaughton et al., 2008; Mineur et al., 2006). FMR1 KO mice also demonstrated increased seizure susceptibility and anxiety – two areas also associated with fragile-X and ASD (El Idrissi et al., 2005; McNaughton et al., 2008; Musumeci et al., 2007; Westmark et al., 2009).

Increased mGluR5 activity in the FMR1^{−/−} mouse led to the prediction that mGluR5 antagonist may rescue some of these phenotypes. Indeed, chronic treatment with the mGluR5 antagonist 2-methyl-6-(phenylethynyl)-pyridine (MPEP), can rescue morphological and synaptic phenotypes, while reducing anxiety and seizure phenotypes, yet the core ASD features comprising social deficits are resistant to MPEP rescue (de Vrij et al., 2008; Meredith et al., 2010; Suvrathan et al., 2010; Yan et al., 2005). In contrast, FMR1 rescue itself does reverse social deficits (Spencer et al., 2008). These data first established a theme that glutamatergic dysfunction may primarily impact specific symptom domains within the core- and associated-ASD features. These include anxiety, seizure susceptibility and, as we will discuss below, repetitive behaviors. Additional evidence indicates that chronic MPEP treatment efficacy in FMR1 mice models is developmentally dependent, suggesting that glutamatergic dysfunction has a role in the developmental etiology of fragile-X (Meredith et al., 2010). While mGluR5 antagonists do not appear to be a magic bullet for fragile-X or ASD treatment, their ability to combat anxiety and reduce the increased risk for epilepsy associated with these disorders, may be clinically significant in treating ASD.

4. Indirect impact on glutamatergic system

4.1. Disruption of synaptic proteins in excitatory synapses

In the case of GluR6 and FMR1 abnormalities, glutamate receptor disruptions are directly implicated. An indirect, but strong case for examining glutamate receptors and synapses in models of ASD is the gathering evidence for disruption synapse development and homeostasis in ASD. (Betancur et al., 2009; Zoghbi, 2003). This evidence is supported by studies linking a large family of candidate genes to ASD, which are involved in the plasticity, maintenance, and development of excitatory synapses. These candidate genes include the presynaptic factors neuroligin-1 as well as neuregulins-1, 3 and 4; members of the trans-synaptic matrix, neural cell adhesion molecule-2 (NCAM2); and, post-synaptically, neuroligins, the synaptic-scaffolding proteins (Shank1-3), as well as the post-synaptic density protein, PSD95 (Berkel et al.; Betancur et al., 2009; Blundell et al., 2010; Etherton et al., 2009; Feyder et al., 2010). Additionally, the VPA, MeCP2^{−/−}, and fragile-X models demonstrate disruption of synaptic ASD candidate proteins such as neuroligin 3 (Dahlhaus and El-Husseini, 2010; Gandal et al., 2010; Kolozi et al., 2009). When these synaptic proteins are transgenically disrupted in mice, the most common behavioral impact is increased stereotypies and signs of anxiety, though in some models social deficits are also found (see: Table 1; Blundell et al., 2010; Etherton et al., 2009; Feyder et al., 2010). Probing synaptic transmission in these models also often implicates reduced AMPA- and NMDA-dependent glutamatergic transmission. As in fragile-X, there is also pharmacological evidence for treating some of the behavioral outcomes found in models of these protein-specific dysfunctions with mGluR5 antagonists.

4.2. mGluR5

The potential clinical efficacy of NMDA and mGluR5 agents in ASD and PPDs arises from the evidence that disruption of glutamatergic transmission may be a general mechanism underlying at least some of the symptoms in ASD (Silverman et al., 2010a). Because mGluR5 is linked with synaptic plasticity and is critical to the appropriate development and maintenance of excitatory synapses in the CNS, it is a reasonable candidate (Simonyi et al., 2005; Spires et al., 2005). Following this line of reasoning, Jackie Crawley's group tested MPEP in the BTBR mouse (Silverman et al., 2010a). The BTBR mouse is a model of insulin resistance that has demonstrated strong face validity for ASD core symptoms of repetitive behaviors and reduced sociability

Table 1
Selected ASD mouse models that demonstrate a role for glutamate receptors. These models include four examples of disrupted synaptic adhesion or scaffolding proteins. Themes of disrupted excitatory transmission, linked with repetitive behavior and increased anxiety are common among these models. Social deficits associated with these models appear resistant to glutamate receptor agents.

ASD Model	Construct validity	Face validity: ASD core domains	Face validity: ASD-associated symptoms domains	Evidence for disrupted glutamate receptor activity	References
GluR6 ^{-/-} (GRIK2)	Candidate gene		Decreased anxiety, reduced seizure susceptibility	KO directly removes a primary subunit of Kainate receptor type 2.	(Fisahn et al., 2005; Malkesman et al., 2010; Shaltiel et al., 2008)
Fmr1 ^{-/-}	Fmr1 expression is disrupted in fragile-X.	Social deficits	Increase anxiety, and seizure susceptibility	mGluR5 activity increased. Reduced NMDA receptor expression. Disrupted AMPA receptor cycling. mGluR5 antagonist, and mGluR genetic reduction, reduce anxiety and seizure susceptibility.	(Eadie et al., 2010; Nakamoto et al., 2007; Westmark et al., 2009)
BTBR mouse		Social deficits, repetitive behavior		Disrupted metabolism of the endogenous glutamate antagonist kynurenic acid MPEP efficacy in reducing stereotypies.	(McFarlane et al., 2008; Moy et al., 2008; Silverman et al., 2010a)
Prenatal valproic acid exposure (VPA model)	Analogous to prenatal VPA in humans	Social deficits	Gamma-band EEG phenotypes associated with ASD	mGluR5 up-regulation MPEP efficacy Increased NMDA activity.	(Gandal et al., 2010; Rinaldi et al., 2007; Schneider and Przewlocki, 2005)
MECP2 ^{-/-}	Rett syndrome models, MeCP2 expression reduced in ASD. Candidate gene	Social deficits, repetitive behavior		Decreased excitatory transmission. Reduced NMDA expression.	(Chen et al., 2001; Guy et al., 2001; Maliszewska-Cyna et al., 2010; Santos et al., 2007)
SHANK1 ^{-/-}	Candidate gene	Social deficits	Increased anxiety	Excitatory synapse disruption. Reduced Excitatory transmission.	(Hung et al., 2008; Silverman et al., 2010b)
SHANK3 ^{-/+}	Candidate gene	Social deficits		Excitatory synapse disruption. Reduced Excitatory transmission.	Bozdagi et al. (2010)
DLG-4 ^{-/-} (PSD95 analogue)	Candidate gene	Social deficits	Increased anxiety	Excitatory synapse disruption. MPEP reduces anxiety.	Feyder et al. (2010)
Neurexin-1 α ^{-/-}	Candidate gene	Repetitive behavior	Cognitive deficits	Reduced excitatory transmission	Etherton et al. (2009)
Neurologin-1 ^{-/-}	Candidate gene	Repetitive behavior	Cognitive deficits	Reduced NMDA/AMPA ratio	Blundell et al. (2010)

(Moy et al., 2008). Treatment with the mGluR5 antagonist MPEP successfully reversed the repetitive behaviors, but did not increase sociability. This study suggested a limited, but nevertheless promising, role for mGluR5 antagonists in treating core symptoms of ASD (Silverman et al., 2010a). Importantly, this work also indicated that MPEP and other mGluR antagonists are useful as a provisional test of predictive validity in models of ASD.

Similarly, in a mouse model where the homologue for human post-synaptic density protein 95 (PSD-95; mouse DLG-4) is disrupted, the mGluR5 antagonist MPEP was able to reduce signs of anxiety, but social deficits found in these mice were unaffected (Feyder et al., 2010). These mGluR5 findings may be directly related to rescuing specific synaptic abnormalities in the DLG-4 KO mice, as mGluR5, NMDA receptors, and ASD candidate genes in the SHANK family are functionally and physically associated with PSD-95 in the post-synaptic membrane (Guo et al., 2004; Tu et al., 1999).

Recent studies of MPEP in the Valproic acid (VPA) model of prenatally induced ASD further indicated a role for mGluR5. Mice prenatally exposed to VPA developed a number of core symptoms of ASD, paralleling earlier work in rats (Schneider and Przewlocki, 2005). Prenatal VPA exposure in mice also generates mice with intermediate EEG phenotypes of ASD (Gandal et al., 2010). The EEG phenotypes exhibited abnormal Gamma oscillations, which were reversed in the presence of MPEP (Gandal et al., 2010). Gamma abnormalities in response to auditory stimulation are an endophenotype of ASD (Rojas et al., 2008; Wilson et al., 2007). EEG abnormalities are an attractive endpoint, as they can be an intermediate endophenotype shared between models and patients, providing an acute readout of drug effects. Furthermore disrupted gamma-band activity signals a disruption of the circuit machinery generating gamma oscillations (Fries et

al., 2007; Uhlhaas and Singer, 2010). Thus, these results provide a template for using EEG-intermediate phenotypes along with behavioral outcome measures for studying the role of glutamatergic agents in preclinical models of ASD.

4.3. NMDA receptors

In a number of models of ASD, NMDA receptors appear disrupted (Blundell et al., 2010; Eadie et al., 2010; Maliszewska-Cyna et al., 2010; Moy et al., 2008; Rinaldi et al., 2007). Clinically, there have been suggestions that the weak NMDA antagonists, amantadine, memantine and dextromethorphan, may have effectiveness in treating ASD (Chez et al., 2007; Erickson et al., 2007; King et al., 2001; Niederhofer, 2007). These clinical results would predict that NMDA hyperfunction has a role in ASD. NMDA receptor increases have also been found in a rat prenatal-VPA model (Rinaldi et al., 2007). In contrast, NMDA receptor levels are decreased in the FMRP1^{-/-} mouse (Eadie et al., 2010), and in the MeCP2^{-/-} mouse model of RTT (Maliszewska-Cyna et al., 2010). Targeting NMDA receptors in some models has been effective in normalizing behavior. In Neuregulin-1 KO mice, use of the NMDA coagonist D-cycloserine reduced potential signs of anxiety and repetitive behavior, indicating that increasing NMDA activity had a role to play in some ASD-associated symptoms (Blundell et al., 2010).

This lack of concurrence among models and the modest indications of NMDA hyperfunction in ASD suggest that NMDA differences may not be strongly linked to etiologies of ASD. Alternatively, these findings could be ascribed to a failure of the discussed models to replicate NMDA dysfunction as it occurs in ASD. Whatever the explanation in the context of hyperfunction, models of NMDA

hypofunction, which may have some validity for schizophrenia, can also have strong face validity for social deficits in ASD, and may be helpful in determining a role for NMDA in generating autistic phenotypes (Duncan et al., 2004; Moy et al., 2008; Ramsey, 2009).

4.4. AMPA receptors

The reduction of plasticity and post-synaptic excitatory potentials (EPSPs) in many models of ASD indicates downregulation of AMPA receptors in these models (as has been found in ASD (Purcell et al., 2001)). Long-term potentiation (LTP) and depression (LTD) are also often mediated by changes in AMPA receptor subunit expression in the post-synaptic membrane. These forms of synaptic plasticity are strongly associated with learning and memory, and disruption of these may underlie intellectual disabilities in ASD. AMPA receptors have been targeted in human patients using the nootropic piracetam, which works as an allosteric agonist at AMPA receptors, and has met with some potential success as an adjunct therapy (Akhondzadeh et al., 2008). It should be noted, however, that piracetam has many other potential targets in addition to AMPA receptors (Winblad, 2005). Ampakines, which also target AMPA channels, have been successfully used in treating some non-ASD-associated symptoms in the MeCP2^{-/-} mouse models. In the case of ampakines, efficacy is attributed primarily to increased BDNF release (Lauterborn et al., 2000). Because targeting AMPA could provide for rescue of excitatory transmitter deficits, facilitation of LTP or LTD, recovery of cognitive deficits, and could also serve as an indirect means of increasing neurotrophin activity, there is a strong rationale for such work in models of ASD even without current clinical success.

5. Conclusion

5.1. Glutamate system dysfunction is a limited but common theme in models of ASD

Data from these ASD models indicate that excitatory transmission is often disrupted in models of ASD. Current work demonstrates that specific disruption of one candidate gene, or, in the case of VPA, models of induced ASD phenotypes, leads to disruption of other known candidate genes. Thus, these genes can be thought to form networks, whose interactions are important for normal synaptic function. Pharmacologically or genetically targeting mGluRs, GluR6, and NMDA, as well as a number of ASD-candidate genes, suggests that glutamatergic-associated changes within these networks may form mechanistic and behavioral subsets within a broader set of etiologies and phenotypes that co-occur in ASD. At the present juncture, this subset appears to link disrupted excitatory transmission to increased seizure susceptibility, anxiety, and the ASD-core symptom of repetitive behaviors and interests (Table 1). There is much less evidence that glutamatergic dysfunction mediates social deficits. Indeed, FMR1^{-/-} mice demonstrated social deficits that are not mediated by mGluR5, suggesting a discrete non-glutamatergic mechanistic domain underlying social deficits in fragile-X. By contrast, neuroligin expression appears to interact with social deficits in the FMR1^{-/-} model (Dahlhaus and El-Husseini, 2010), suggesting that FMR1 KO precipitates at least two distinct sets of impairments, only one of which is explained by the mGluR5 hypothesis.

The current work indicates that glutamate dysfunction, in the form of reduced excitatory transmission, dysfunction of NMDA mediated plasticity, and mGluR mediated signal transduction, has a role to play in the cognitive and behavioral pathologies of ASD. This hypothesis, delineating a role for disrupted glutamate signaling in ASD, needs to be more fully tested. To do so, studies similar to those described in the FMR1^{-/-} mouse could be performed in a broad range of ASD models to determine if glutamate dysfunction mediates specific behaviors common to ASD models.

In transgenic and in models of ASD (including those that are possibly induced), the impact on glutamate receptors is primarily indirect. Thus the current models have a latent utility for exploring how different genetic risk factors interact to disrupt glutamate transmission, and how disrupted glutamate transmission may interact with other ASD-associated neuropathologies to generate the full range of ASD-associated behaviors. To directly determine the role of glutamate dysfunction on ASD-related behaviors glutamate receptors need to be specifically targeted for hyper- or hypofunction. Such models may be more tractable for preclinical development and tuning of therapeutics to treat glutamate receptor dysfunction in the autistic brain.

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References

- Akhondzadeh S, Tajdar H, Mohammadi MR, Mohammadi M, Nouroozinejad GH, Shabstari OL, et al. A double-blind placebo controlled trial of piracetam added to risperidone in patients with autistic disorder. *Child Psychiatry Hum Dev* 2008;39: 237–45.
- Bardoni B, Mandel JL. Advances in understanding of fragile X pathogenesis and FMRP function, and in identification of X linked mental retardation genes. *Curr Opin Genet Dev* 2002;12:284–93.
- Bear MF, Huber KM, Warren ST. The mGluR theory of fragile X mental retardation. *Trends Neurosci* 2004;27:370–7.
- Berkel S, Marshall CR, Weiss B, Howe J, Roeth R, Moog U, et al. Mutations in the SHANK2 synaptic scaffolding gene in autism spectrum disorder and mental retardation. *Nat Genet* 2010;42:489–91.
- Betancur C, Sakurai T, Buxbaum JD. The emerging role of synaptic cell-adhesion pathways in the pathogenesis of autism spectrum disorders. *Trends Neurosci* 2009;32:402–12.
- Blundell J, Blaiss CA, Etherton MR, Espinosa F, Tabuchi K, Walz C, et al. Neuroligin-1 deletion results in impaired spatial memory and increased repetitive behavior. *J Neurosci* 2010;30:2115–29.
- Bozdagi O, Sakurai T, Papapetrou D, Wang X, Dickstein DL, Takahashi N, et al. Haploinsufficiency of the autism-associated Shank3 gene leads to deficits in synaptic function, social interaction, and social communication. *Mol Autism* 2010;1:15.
- Brodtkin ES. Social behavior phenotypes in fragile X syndrome, autism, and the Fmr1 knockout mouse: theoretical comment on McNaughton et al. (2008). *Behav Neurosci* 2008;122:483–9.
- Bruneau EG, Esteban JA, Akaaboune M. Receptor-associated proteins and synaptic plasticity. *FASEB J* 2009;23:679–88.
- Chen RZ, Akbarian S, Tudor M, Jaenisch R. Deficiency of methyl-CpG binding protein-2 in CNS neurons results in a Rett-like phenotype in mice. *Nat Genet* 2001;27: 327–31.
- Chez MG, Burton Q, Dowling T, Chang M, Khanna P, Kramer C. Memantine as adjunctive therapy in children diagnosed with autistic spectrum disorders: an observation of initial clinical response and maintenance tolerability. *J Child Neurol* 2007;22: 574–9.
- Crawley JN. Mouse behavioral assays relevant to the symptoms of autism. *Brain Pathol* 2007;17:448–59.
- Dahlhaus R, El-Husseini A. Altered neuroligin expression is involved in social deficits in a mouse model of the fragile X syndrome. *Behav Brain Res* 2010;208:96–105.
- de Vrij FM, Levenga J, van der Linde HC, Koekkoek SK, De Zeeuw CI, Nelson DL, et al. Rescue of behavioral phenotype and neuronal protrusion morphology in Fmr1 KO mice. *Neurobiol Dis* 2008;31:127–32.
- den Broeder MJ, van der Linde H, Brouwer JR, Oostra BA, Willemsen R, Ketting RF. Generation and characterization of FMR1 knockout zebrafish. *PLoS ONE* 2009;4: e7910.
- Dolen G, Bear MF. Role for metabotropic glutamate receptor 5 (mGluR5) in the pathogenesis of fragile X syndrome. *J Physiol* 2008;586:1503–8.
- Dolen G, Carpenter RL, Ocain TD, Bear MF. Mechanism-based approaches to treating fragile X. *Pharmacol Ther* 2010;127:78–93.
- Duncan GE, Moy SS, Perez A, Eddy DM, Zinzow WM, Lieberman JA, et al. Deficits in sensorimotor gating and tests of social behavior in a genetic model of reduced NMDA receptor function. *Behav Brain Res* 2004;153:507–19.
- Eadie BD, Cushman J, Kannagara TS, Fanselow MS, Christie BR. NMDA receptor hypofunction in the dentate gyrus and impaired context discrimination in adult Fmr1 knockout mice. *Hippocampus* 2010.
- El Idrissi A, Ding XH, Scalia J, Trenkner E, Brown WT, Dobkin C. Decreased GABA(A) receptor expression in the seizure-prone fragile X mouse. *Neurosci Lett* 2005;377: 141–6.

- Erickson CA, Posey DJ, Stigler KA, Mullett J, Katschke AR, McDougle CJ. A retrospective study of memantine in children and adolescents with pervasive developmental disorders. *Psychopharmacology* 2007;191:141–7.
- Etherton MR, Blaiss CA, Powell CM, Sudhof TC. Mouse neuroligin-1 alpha deletion causes correlated electrophysiological and behavioral changes consistent with cognitive impairments. *Proc Natl Acad Sci USA* 2009;106:17998–8003.
- Feyder M, Karlsson RM, Mathur P, Lyman M, Bock R, Momenan R, et al. Association of mouse *Dlg4* (PSD-95) gene deletion and human *DLG4* gene variation with phenotypes relevant to autism spectrum disorders and Williams' Syndrome. *Am J Psychiatry* 2010;167:1508–17.
- Fisahn A, Contractor A, Traub RD, Buhl EH, Heinemann SF, McBain CJ. Distinct roles for the kainate receptor subunits GluR5 and GluR6 in kainate-induced hippocampal gamma oscillations. *J Neurosci* 2004;24:9658–68.
- Fisahn A, Heinemann SF, McBain CJ. The kainate receptor subunit GluR6 mediates metabotropic regulation of the slow and medium AHP currents in mouse hippocampal neurones. *J Physiol* 2005;562:199–203.
- Fries P, Nikolic D, Singer W. The gamma cycle. *Trends Neurosci* 2007;30:309–16.
- Gandal MJ, Edgar JC, Ehrlichman RS, Mehta M, Roberts TP, Siegel SJ. Validating gamma oscillations and delayed auditory responses as translational biomarkers of autism. *Biol Psychiatry* 2010;68:1100–6.
- Guo W, Wei F, Zou S, Robbins MT, Sugiyo S, Ikeda T, et al. Group I metabotropic glutamate receptor NMDA receptor coupling and signaling cascade mediate spinal dorsal horn NMDA receptor 2B tyrosine phosphorylation associated with inflammatory hyperalgesia. *J Neurosci* 2004;24:9161–73.
- Guy J, Hendrich B, Holmes M, Martin JE, Bird A. A mouse *Mecp2*-null mutation causes neurological symptoms that mimic Rett syndrome. *Nat Genet* 2001;27:322–6.
- Holt R, Barnby G, Maestrini E, Bacchelli E, Brocklebank D, Sousa I, et al. Linkage and candidate gene studies of autism spectrum disorders in European populations. *Eur J Hum Genet* 2010;18:1013–9.
- Hung AY, Futai K, Sala C, Valtchanoff JG, Ryu J, Woodworth MA, et al. Smaller dendritic spines, weaker synaptic transmission, but enhanced spatial learning in mice lacking Shank1. *J Neurosci* 2008;28:1697–708.
- Jamain S, Betancur C, Quach H, Philippe A, Fellous M, Giros B, et al. Linkage and association of the glutamate receptor 6 gene with autism. *Mol Psychiatry* 2002;7:302–10.
- Kaytor MD, Orr HT. RNA targets of the fragile X protein. *Cell* 2001;107:555–7.
- Kim SA, Kim JH, Park M, Cho IH, Yoo HJ. Family-based association study between *GRIK2* polymorphisms and autism spectrum disorders in the Korean trios. *Neurosci Res* 2007;58:332–5.
- King BH, Wright DM, Handen BL, Sikich L, Zimmerman AW, McMahon W, et al. Double-blind, placebo-controlled study of amantadine hydrochloride in the treatment of children with autistic disorder. *J Am Acad Child Adolesc Psychiatry* 2001;40:658–65.
- Kolozsi E, Mackenzie RN, Roulet FI, deCatanzaro D, Foster JA. Prenatal exposure to valproic acid leads to reduced expression of synaptic adhesion molecule neuroigin 3 in mice. *Neuroscience* 2009;163:1201–10.
- Korff S, Harvey BH. Animal models of obsessive-compulsive disorder: rationale to understanding psychobiology and pharmacology. *Psychiatr Clin North Am* 2006;29:371–90.
- Kullmann DM, Lamsa K. Roles of distinct glutamate receptors in induction of anti-Hebbian long-term potentiation. *J Physiol* 2008;586:1481–6.
- Lauterborn JC, Lynch G, Vanderklish P, Arai A, Gall CM. Positive modulation of AMPA receptors increases neurotrophin expression by hippocampal and cortical neurons. *J Neurosci* 2000;20:8–21.
- Lodge D. The history of the pharmacology and cloning of ionotropic glutamate receptors and the development of idiosyncratic nomenclature. *Neuropharmacology* 2009;56:6–21.
- Maliszewska-Cyna E, Bawa D, Eubanks JH. Diminished prevalence but preserved synaptic distribution of N-methyl-D-aspartate receptor subunits in the methyl CpG binding protein 2 (*Mecp2*)-null mouse brain. *Neuroscience* 2010;168:624–32.
- Malkesman O, Scattoni ML, Paredes D, Tragon T, Pearson B, Shaltiel G, et al. The female urine sniffing test: a novel approach for assessing reward-seeking behavior in rodents. *Biol Psychiatry* 2010;67:864–71.
- McBride SM, Choi CH, Wang Y, Liebelt D, Braunstein E, Ferreiro D, et al. Pharmacological rescue of synaptic plasticity, courtship behavior, and mushroom body defects in a *Drosophila* model of fragile X syndrome. *Neuron* 2005;45:753–64.
- McFarlane HG, Kusek GK, Yang M, Phoenix JL, Bolivar VJ, Crawley JN. Autism-like behavioral phenotypes in BTBR T + tf/j mice. *Genes Brain Behav* 2008;7:152–63.
- McNaughton CH, Moon J, Strawderman MS, Maclean KN, Evans J, Strupp BJ. Evidence for social anxiety and impaired social cognition in a mouse model of fragile X syndrome. *Behav Neurosci* 2008;122:293–300.
- Meredith RM, de Jong R, Mansvelter HD. Functional rescue of excitatory synaptic transmission in the developing hippocampus in *Fmr1*-KO mouse. *Neurobiol Dis* 2010;41:104–10.
- Mineur YS, Huynh LX, Crusio WE. Social behavior deficits in the *Fmr1* mutant mouse. *Behav Brain Res* 2006;168:172–5.
- Morales J, Hiesinger PR, Schroeder AJ, Kume K, Verstreken P, Jackson FR, et al. *Drosophila* fragile X protein, DFXR, regulates neuronal morphology and function in the brain. *Neuron* 2002;34:961–72.
- Moy SS, Nadler JJ, Poe MD, Nonneman RJ, Young NB, Koller BH, et al. Development of a mouse test for repetitive, restricted behaviors: relevance to autism. *Behav Brain Res* 2008;188:178–94.
- Musumeci SA, Calabrese G, Bonaccorso CM, D'Antoni S, Brouwer JR, Bakker CE, et al. Audiogenic seizure susceptibility is reduced in fragile X knockout mice after introduction of *FMR1* transgenes. *Exp Neurol* 2007;203:233–40.
- Nakamoto M, Nalavadi V, Epstein MP, Narayanan U, Bassell GJ, Warren ST. Fragile X mental retardation protein deficiency leads to excessive mGluR5-dependent internalization of AMPA receptors. *Proc Natl Acad Sci USA* 2007;104:15537–42.
- Niederhofer H. Glutamate antagonists seem to be slightly effective in psychopharmacologic treatment of autism. *J Clin Psychopharmacol* 2007;27:317–8.
- Orehkova EV, Stroganova TA, Nygren G, Tsetlin MM, Posikera IN, Gillberg C, et al. Excess of high frequency electroencephalogram oscillations in boys with autism. *Biol Psychiatry* 2007;62:1022–9.
- Polleux F, Lauder JM. Toward a developmental neurobiology of autism. *Ment Retard Dev Disabil Res Rev* 2004;10:303–17.
- Purcell AE, Jeon OH, Zimmerman AW, Blue ME, Pevsner J. Postmortem brain abnormalities of the glutamate neurotransmitter system in autism. *Neurology* 2001;57:1618–28.
- Ramsey AJ. NR1 knockdown mice as a representative model of the glutamate hypothesis of schizophrenia. *Prog Brain Res* 2009;179:51–8.
- Rinaldi T, Kulangara K, Antonello K, Markram H. Elevated NMDA receptor levels and enhanced postsynaptic long-term potentiation induced by prenatal exposure to valproic acid. *Proc Natl Acad Sci USA* 2007;104:13501–6.
- Rodriguez-Moreno A, Sihra TS. Metabotropic actions of kainate receptors in the CNS. *J Neurochem* 2007;103:2121–35.
- Rojas DC, Maharajh K, Teale P, Rogers SJ. Reduced neural synchronization of gamma-band MEG oscillations in first-degree relatives of children with autism. *BMC Psychiatry* 2008;8:66.
- Santos M, Silva-Fernandes A, Oliveira P, Sousa N, Maciel P. Evidence for abnormal early development in a mouse model of Rett syndrome. *Genes Brain Behav* 2007;6:277–86.
- Schneider T, Przewlocki R. Behavioral alterations in rats prenatally exposed to valproic acid: animal model of autism. *Neuropsychopharmacology* 2005;30:80–9.
- Shaltiel G, Maeng S, Malkesman O, Pearson B, Schloesser RJ, Tragon T, et al. Evidence for the involvement of the kainate receptor subunit GluR6 (*GRIK2*) in mediating behavioral displays related to behavioral symptoms of mania. *Mol Psychiatry* 2008;13:858–72.
- Shuang M, Liu J, Jia MX, Yang JZ, Wu SP, Gong XH, et al. Family-based association study between autism and glutamate receptor 6 gene in Chinese Han trios. *Am J Med Genet B Neuropsychiatr Genet* 2004;131B:48–50.
- Silverman JL, Tolu SS, Barkan CL, Crawley JN. Repetitive self-grooming behavior in the BTBR mouse model of autism is blocked by the mGluR5 antagonist MPEP. *Neuropsychopharmacology* 2010a;35:976–89.
- Silverman JL, Turner SM, Barkan CL, Tolu SS, Saxena R, Hung AY, et al. Sociability and motor functions in Shank1 mutant mice. *Brain Res* 2010b.
- Simonyi A, Schachtman TR, Christoffersen GR. The role of metabotropic glutamate receptor 5 in learning and memory processes. *Drug News Perspect* 2005;18:353–61.
- Spencer CM, Graham DF, Yuva-Paylor LA, Nelson DL, Paylor R. Social behavior in *Fmr1* knockout mice carrying a human *FMR1* transgene. *Behav Neurosci* 2008;122:710–5.
- Spires TL, Molnar Z, Kind PC, Cordery PM, Upton AL, Blakemore C, et al. Activity-dependent regulation of synapse and dendritic spine morphology in developing barrel cortex requires phospholipase C-beta1 signalling. *Cereb Cortex* 2005;15:385–93.
- Strutz-Seebohm N, Korniyuchuk G, Schwarz R, Baltaev R, Ureche ON, Mack AF, et al. Functional significance of the kainate receptor GluR6 (*M836I*) mutation that is linked to autism. *Cell Physiol Biochem* 2006;18:287–94.
- Suvrathan A, Hoeffler CA, Wong H, Klann E, Chattarji S. Characterization and reversal of synaptic defects in the amygdala in a mouse model of fragile X syndrome. *Proc Natl Acad Sci USA* 2010;107:11591–6.
- Svitkina T, Lin WH, Webb DJ, Yasuda R, Wayman GA, Van Aelst L, Soderling SH. Regulation of the postsynaptic cytoskeleton: roles in development, plasticity, and disorders. *J Neurosci* 2010;30:14937–42.
- Tu JC, Xiao B, Naisbitt S, Yuan JP, Petralia RS, Brakeman P, et al. Coupling of mGluR/Homer and PSD-95 complexes by the Shank family of postsynaptic density proteins. *Neuron* 1999;23:583–92.
- Tucker B, Richards RI, Lardelli M. Contribution of mGluR and *Fmr1* functional pathways to neurite morphogenesis, craniofacial development and fragile X syndrome. *Hum Mol Genet* 2006;15:3446–58.
- Uhlhaas PJ, Singer W. Abnormal neural oscillations and synchrony in schizophrenia. *Nat Rev* 2010;11:100–13.
- Wan L, Dockendorff TC, Jongens TA, Dreyfuss G. Characterization of dFMR1, a *Drosophila* melanogaster homolog of the fragile X mental retardation protein. *Mol Cell Biol* 2000;20:8536–47.
- Westmark CJ, Westmark PR, Malter JS. MPEP reduces seizure severity in *Fmr1* KO mice over expressing human Abeta. *Int J Clin Exp Pathol* 2009;3:56–68.
- Wilson TW, Rojas DC, Reite ML, Teale PL, Rogers SJ. Children and adolescents with autism exhibit reduced MEG steady-state gamma responses. *Biol Psychiatry* 2007;62:192–7.
- Winblad B. Piracetam: a review of pharmacological properties and clinical uses. *CNS Drug Rev* 2005;11:169–82.
- Yan QJ, Mammal M, Tranfaglia M, Bauchwitz RP. Suppression of two major fragile X syndrome mouse model phenotypes by the mGluR5 antagonist MPEP. *Neuropharmacology* 2005;49:1053–66.
- Zoghbi HY. Postnatal neurodevelopmental disorders: meeting at the synapse? *Science* 2003;302:826–30.